A Thermal Degradation (Thermolysis) Study of Rotenone Extracted from *Derris elliptica* Roots Using Reverse-Phase High Performance Liquid Chromatography (RP-HPLC)

(Kajian Degradasi Terma (Termolisis) bagi Rotenon dalam Ceaicr Ekstrak Mentah Pekat (CLCE) Diekstrak daripada Akar *Derris elliptica* Menggunakan Fasa Keterbalikan Ceaicr Kromatografi Berprestasi Tinggi (HPLC))

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ABSTRACT

Bio-pesticides are becoming increasingly important as pest management tools in various cropping systems in the tropics essentially to remedy problems associated with the indiscriminate use of ‘hard’ and non-environmental friendly inorganic pesticide. In these past few decades, many bio-pesticidal products, both microbial-based (bacteria, fungi, microspora, entomopathogenic nematodes and viruses) and plant-based botanicals (rotenone and azadiracthin) have been studied for their use against insect pests in the tropics. In this study, the effects of the concentration process with respect to the yield of rotenone (mg) and its concentration (mg/mL) are presented extensively. The raw plants were collected from Kota Johor Lama, Johor and sorted to obtain the roots and stems. Only the roots and stems were utilized as raw materials of the extraction process. The rotenone from roots and stems was extracted using the normal soaking extraction (NSE) at 28 to 30°C with 95% (v/v) of acetone as a solvent and the solvent-to-solid ratio of 10 mL/g. The extraction was carried out for 24 h. Next, the liquid crude extract was concentrated using the rotary evaporator at 50°C and 80 mbar of vacuum pressure to remove approximately 90% of solvent. The fractions of the liquid crude extract were collected (15 min/mL fraction), diluted (1/100 with acetone) and cleaned up (to remove any fine debris) prior to determination of rotenone content (mg) and concentration (mg/mL) by using the reverse-phase high performance liquid chromatography (RP-HPLC).

Finally, the results showed that there was a significant effect of thermal degradation or dissipation of rotenone content at higher operating temperature (greater than 40°C) with a rapid rotenone reduction for the first 15 min of exposure. The possibilities for better exploitation and identification of the effective operating parameters based on the above mentioned results will be perhaps discussed in the future.

Keywords: Concentrated liquid crude extract; *Derris elliptica*; rotenone; thermal degradation

ABSTRAK

Bio-pestisid pada masa ini sedang mengalami peningkatan yang sangat penting sebagai kaedah mengawal makhluk perosak dalam variasi sistem tanaman tropika terutamanya sebagai jalan untuk mengatasi masalah yang berkaitan dengan penyalahgunaan pestisid bahan bukan organik berat yang tidak mesra alam. Dalam beberapa dekad yang lalu, banyak produk bio-pestisid, kedua-duanya bagi basis mikrob (bakteria, kulat, mikrosprodia, entomopatogenik nematod dan virus) dan bahan botani berasaskan tumbuhan (rotenon dan azadiraktin) telah dikaji berkesan untuk kegunaan serangan perosak dalam iklim tropika. Dalam kajian ini, khas daripada proses pemekatan terhadap keberhasilan rotenon (mg) dan kepekatannya (mg/mL) telah dikaji secara menyeluruh. Sumber pokok mentah dikumpulkan dari Kota Johor Lama, Johor dan diasingkan untuk pengumpulan akar dan batang. Hanya akar dan batang sahaja digunakan sebagai bahan mentah untuk proses pengekstrakan. Rotenon daripada bagian akar dan batang diekstrak menggunakan pengekstrakan celuran norma pada suhu bulik iaitu 28 to 30°C dengan 95% (v/v) aseton sebagai pelarut dan nisbah pelarut-kepadak-pepejal bagi proses pengekstrakan adalah 10 mL/g. Proses pengekstrakan dijalankan selama 1440 min. Seterusnya, ceaicr ekstrak mentah dipekatkan menggunakan alat pengekstrakan berpusing pada suhu 50°C dan tekanan vakum 80 mbar untuk mengeluarkan pelarut aseton sebanyak kurang 90%. Fraksi ceaicr ekstrak mentah dikumpulakan (15 min/mL fraksi), dicairkan (1/100 dengan aseton) dan ditapis (untuk membuang sebarang habuk halus) sebelum proses analisis kandungan rotenon (mg) dan kepekatannya (mg/mL) menggunakan fasa terbalik ceaicr kromatografi berprestasi tinggi (HPLC). Akhirnya, keputusan uji kajian menunjukkan bahawa terdapat kesan signifikan degradasi terma terhadap kandungan rotenon pada suhu operasi tinggi (melebihi 40°C) dengan kadar pengurangan pantas rotenon pada 15 min pertama operasi. Kebarangkalian untuk mengkaji dan mengenal pasti parameter operasi yang lebih efektif berdasarkan kepada keputusan uji kajian ini akan dibincangkan kelak.

Kata kunci: Ceaicr ekstrak mentah pekat; degradasi terma; *Derris elliptica*; rotenone
INTRODUCTION

*Derris elliptica* or ‘Tuba’ as it is known locally is an insecticidal plant in Malaysia that has been used for the purpose of bio-pesticide production. ‘Tuba’ plant is a kind of woody creeper plant and a climber. It needs at least 75% soil moisture content and the surrounding temperature should be 25 to 30°C to obtain high content of rotenone during its development. A calm area with low acidity of soil content will enhance the production of rotenone (Grinda & Gueyne 1986). ‘Tuba’ is a member of the Leguminosae, Fabaceae family, which comprises of 200 genera and 68 species including 21 species of Tephrosia, 12 of *Derris*, 12 of Lonchocarpus, 10 of Millettia and several of Mundula (John & Ron 1944). Three species are found in Malaysia, which are *Derris elliptica*, *Derris malaccensis* and *Derris uliginosa* (Gaby 1986). *Derris* is a climbing plant in Southeast Asia and its roots contain rotenone which is derived from one of the secondary metabolites in isoflavonoids group (Figure 1). The Amazonian *Derris elliptica* and *Derris malaccensis* contain approximately 4 to 5% (w/w) rotenone while *Lonchocarpus utilis* and *Lonchocarpus urucu* contain 8 to 10% (w/w) rotenone in dried roots (Kole et al. 1992). However, rotenone content in the extracts of *Derris elliptica* dried roots collected from the state of Johor, Malaysia has less than 1.2% (w/w) (Zubairi et al. 2014a). Irrespective of its availability in nature, this strong insecticide is commonly used by the aborigines/natives to paralyze the fish and to kill insects that infested their vegetables and fruits (John & Ron 1944). There are many uses of these insecticides that have been discovered. In addition to their effectiveness for both piercing-sucking insects, such as aphids and red bugs and chewing insects, especially caterpillars upon plants, they also make excellent dusts for external parasites of animals such as fleas and lice (Grinda & Gueyne 1986). However, the bio-active constituents (especially rotenone) available in rotenoids resin (Zubairi et al. 2014b, 2014c) are essentially prone to thermal degradation and photochemically unstable (photolysis). The thermal degradation or thermolysis is defined as a chemical decomposition caused by heat/radiation which breaks chemical bonds in the compound (Pielichowski & Njuguna 2008). The breakdown mechanism starts with the energy absorption to break chemical bonds in pesticide molecules which are in the ranges of 70 to 120 kcal mol⁻¹. This energy then excites the molecules and produced oxygen reactive species. The radical molecules (especially rotenone) contained inside the rotenoids resin are believed to deteriorate rapidly into smaller molecules of dihydrorotenone (non-toxic substance) and water when exposed to sunlight and air (surrounding temperature: 35 to 40°C) (Cavoski et al. 2007). Any kind of applications using the extracted rotenoids resin such as spray and dusts typically lose their effectiveness within a week after application (Schnick 1974). For that reason, this study was carried out to observe and to identify the effect of rotenone content extracted from *Derris elliptica* roots and the extent of rotenone dissipation against high operating temperature (greater than 40°C).

**FIGURE 1.** Rotenone molecular structures (Kidd & James 1991; Kole et al. 1992)

MATERIAL AND METHODS

PLANT COLLECTION

*Derris elliptica* was collected in the state of Johor; Kota Johor Lama, Malaysia.

RAW MATERIAL

An important aspect of the phytochemical processing was the pre-processing of the herbal material prior to the extraction process. The treatment of the herbal material affects the viability of the phytochemical as well as the extraction yield (Grinda & Gueyne 1986). The procured *Derris* roots were immediately sent to undergo the cleaning process to remove dirt and soil. The procured *Derris* roots were kept and dried in an oven overnight at room temperature (28 to 30°C) and were sorted to collect the roots and stems. Only roots and stems were utilized. The roots and stems were cut into small pieces prior to grinding.

PREPARATION OF CONCENTRATED LIQUID CRUDE EXTRACT

The extraction process was carried out by soaking 50 g of dried roots and stems in 500 mL of acetone 95% (v/v) with a solvent-to-solid ratio of 10 mL/g for 24 h at room temperature (28 to 30°C). The liquid crude extracts were filtered through a 15 cm Whatman filter paper (number 4) directly into 500 mL beakers after 24 h of extraction. Next, the liquid crude extract was concentrated using the rotary evaporator (Heidolph-Laborata 4001) at 50°C and 80 mbar of vacuum pressure. The fractions of the liquid crude extract were collected interwally (15 min/mL/fraction) and stored in dark vials (2 mL).
ANALYSIS OF THE ROTENONE LIQUID CRUDE EXTRACT

The fractions of the liquid crude extract were subjected to quantitative analysis by using a reverse-phase high performance liquid chromatography (RP-HPLC) with UV (Photodiode Array - PDA) detection at 294 nm to determine the rotenone content and its concentration. Table 1 shows the operating conditions of reversed-phase HPLC isocratic system. The analysis of the extract solutions was carried out by using the external standard method (Rotenone PESTANAL®, analytical grade, 96.2% - Sigma-Aldrich™) as an external standard solution. Table 2 shows the calculation of rotenone yield and actual concentration using RP-HPLC external standard method. The Waters™ Corp. (C18) liquid chromatography stainless steel column with particle size of 10 μm (3.9 mm I.D x 150 mm length) was utilized in the analysis. For the preparation of rotenone standard solution, about 20 mg rotenone standard powder (Rotenone PESTANAL®, analytical grade, 96.2% - Sigma-Aldrich™) was weighed and put into 125 mL Erlenmeyer glass flask. The standard powder was dissolved with 50 mL of analytical grade of acetonitrile; 99.9% (v/v) acetonitrile on a Gyratory shaker for 10 min. After shaking, the standard solution was filtered through a 15 cm Whatman filter paper (number 2) directly into 50 mL beakers. About 10 mL of the standard solution was re-filtered through an organic sample clarification kit (Waters™ Assoc.) containing 0.45/0.5 μm filters to remove impurities during the preparation of the solution. The isocratic solvent system was implemented throughout the whole analysis using acetonitrile and deionized water with a ratio of 60:40 as a mobile phase and the amplitude unit full scale (AUFS) of the detection was 2 (AOAC Official Method 2000; Rodney & Alan 1976).

THIN LAYER CHROMATOGRAPHY

Initially, 1 g of concentrated liquid crude extract (CLCE) and 0.5 mg of rotenone standard powder (Rotenone PESTANAL®, analytical grade, 96.2% - Sigma-Aldrich™) were vortexed and dissolved with chloroform (analytical grade) for 10 to 15 min. Next, the thin layer chromatography (TLC) was carried out to analyze the presence of rotenone and its thermolysis byproduct constituents in the CLCE. Petroleum ether and ethyl acetate with a ratio of 6:4 were used as a mobile phase in the development chamber (Zubairi et al. 2014c). The separation of the bio-active compound was about 30 to 45 min. The markers that appeared onto the TLC silica plate were viewed and illustrated using the ultra-violet (UV) lamp with the wavelength of 254 nm.

STATISTICAL ANALYSIS

Data was presented as mean ± standard deviation (SD) of mean. Statistical comparisons were performed using one-way analysis of variance (ANOVA) for detecting significant differences among sample means of the groups, this was followed by Tukey’s test (PASW version 17.0 IBM Co.) for multiple comparisons to determine values that were significantly different. A \( p<0.05 \) was considered statistically significant.

### TABLE 1. Parameters of RP-HPLC (Baron & Freudenthal 1976)

<table>
<thead>
<tr>
<th>Parameters</th>
<th>Setting</th>
</tr>
</thead>
<tbody>
<tr>
<td>Column temperature (°C)</td>
<td>Ambient</td>
</tr>
<tr>
<td>Flow rate of separation</td>
<td>0.4 mL/min</td>
</tr>
<tr>
<td>UV wavelength (λ)</td>
<td>294 nm</td>
</tr>
<tr>
<td>Injection volume</td>
<td>5 μL</td>
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<tr>
<td>Amplitude unit full scale (AUFS)</td>
<td>2</td>
</tr>
</tbody>
</table>

### TABLE 2. Calculation of rotenone yield and actual concentration using RP-HPLC external standard method

Rotenone standard (Sigma-Aldrich™; purity of 95 - 98% (w/w)):

Rotenone standard concentration \( (C_{std}) = X \) (mg/mL)

Peak area \( (A) = Y \) (mV.s)

Sensitivity Factor \( (SF) = (A)/(C_{std}) = J \) (mV.s. x mL/mg)

Sample concentration (Liquid crude extract (LCE) or concentrated liquid crude extract (CLCE)):

Sample peak area \( (A_{sample}) = Y \) (mV.s)

Sample concentration \( (C_{sample}) = (A_{sample})/SF = Q \) mg/mL

If, the sample involves dilution: (Dilution factor; DF = flask volume/pipette volume)

Actual concentration (rotenone) = \( Q \) mg/mL \( \times DF = G \) (mg/mL)

\[(10 \text{ mg/mL = 1%}) - \text{ To get 1%: } 10/1000 (\text{g/mL}) \times 100\% = 1\%\]

Yield of rotenone = \( C_{sample} \) (mg/mL) \( \times \) volume (mL) of LCE or CLCE = \( (G) \) mg/mL \( \times (mL) = K \) (mg)
RESULTS AND DISCUSSION

The main objective of this study was to observe the thermal degradation profile of rotenone when exposed to high operating temperature (greater than 40°C) as rotenone is commonly known as a light (photolysis) and heat (thermolysis) sensitive compound. Since there is no study conducted to observe the sensitivity of rotenone extracts taken from Malaysia Derris spp. on the thermolysis behaviours, this study was employed. Figure 2 shows that there was a significant reduction of rotenone (mg) for the first 15 min of the concentration process ($p<0.05$). Thus, it appears that rotenone is strongly affected by the temperature greater than 40°C. The initial concentration of the liquid crude extract was observed to be 41.75±3.2 mg/mL (rotenone content: 35,072±155 mg). As the concentration time increased, the amount of rotenone was reduced tremendously by 90±2.5% (w/w) for the first 15 min of the process and the amount of rotenone remained constant until 105 min where the highest concentration occurred (122±5.8 mg/mL: 1,108±146 mg of rotenone) (Figure 3). The reduction of rotenone content in the extract was in line with Grinda and Gueyne’s (1986) findings whereby the rotenone content could only be retained up to 14% (w/w) with the existence of operating temperature up to 45°C for half an hour. Meanwhile, the half-life ($t_{1/2}$) of rotenone under photolysis is predicted to be approximately 3½ h when exposed to bright sunlight (approximately 30 to 40°C) (Grinda & Gueyne 1986; Pagan et al. 1949). For that
CONCLUSION

After some extensive studies have been done, there are several conclusions that can be drawn. First, rotenone is strongly affected by the operating temperature greater than 40°C irrespective of any type of Derris spp. collected (in this study, the roots were collected from the state of Johor, Malaysia). Temperature greater than 40°C will result in dissipation of rotenone into non-toxic compounds. The appearances of several unknown spots (TLC analysis) proved that the existence of rotenone thermolysis byproducts in the concentrated extract were undeniable. Second, to extract the rotenone effectively, the extraction process coupled with sonication is recommended as it ruptures plant cell wall easily and increases the rate of diffusivity into bulk volume of solvent. In fact, it should be done at certain temperature (below than 40°C). Although in principle, as the extraction temperature increases; it will increase the rate of extraction by increasing the internal diffusion as well as the mass transfer coefficient values and thus reducing the extraction time. However, it should be noted that increasing the temperature beyond certain values will lead to a decrease in isoflavonoid compounds’ yield due to the high susceptibility of the isoflavonoid to high temperature.

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