Neuroprotective Effects of Ethyl Acetate Extract of Zingiber zerumbet (L.) Smith against Oxidative Stress on Paraquat-Induced Parkinsonism in Rats (Kesan Neurolindungi Ekstrak Asetat Zingiber zerumbet (L.) Smith terhadap Tekanan Oksidatif pada Tikus Parkinsonisme Aruhan Parakuat)

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ABSTRACT
Zingiber zerumbet has been traditionally used as an anti-inflammation and antioxidant agent. The present study investigates the neuroprotective effects of ethyl acetate extract of Z. zerumbet against oxidative stress on paraquat (PQ)-induced Parkinsonism in rats. Forty male Sprague-Dawley rats were divided into five groups: Negative control (normal saline), positive control (N-acetylcysteine, NAC 20 mg/kg + PQ 10 mg/kg), PQ only, 200 mg/kg Z. zerumbet + PQ and 400 mg/kg Z. zerumbet + PQ. The extract was given orally for 19 consecutive days and PQ was administered intraperitoneally on day 8-12th of the treatment regime. Both serum and fresh brains containing substantia nigra (SN) region were taken for biochemical and histological analysis. Administration of both 200 and 400 mg/kg ethyl acetate Z. zerumbet extracts to the PQ-treated groups have resulted in: Decreased levels of MDA and PC in the SN homogenates; and increased SOD, GPx; and CAT activities in the SN and serum. Overall, ethyl acetate extract of Z. zerumbet reduced oxidative stress in the SN of PQ-induced neuronal damages, therefore, has the potential to be developed as a preventive agent for neurodegenerative disorders caused by environmental toxins.

Keywords: Antioxidant; oxidative stress; paraquat; Parkinson’s disease; Zingiber zerumbet

INTRODUCTION
Parkinson’s disease (PD) is the second most common neurodegenerative disorder after Alzheimer’s disease (Yokohama et al. 2008). Parkinson’s disease exerts an enormous impact on the lives of affected individual, their families and also the society (Giasson et al. 2002). Meta-analysis of 27 international studies showed that PD affects both gender, with increasing number of incidence as age advances. Though the number is higher in males at all age groups, further analysis showed that the incidence affecting the males is significantly higher in the 60-69 and 70-79 age groups compared to the women of the same age groups (Hirsch et al. 2016).

PD is the result of selective loss of dopaminergic neurons in the substantia nigra pars compacta (SNpc) (McCollum et al. 2010). It also involves an abnormal deposition of intracellular inclusions or Lewy bodies within the brain (Lewis & Cookson 2012). Lewy bodies are protein aggregates composed mainly of α-synuclein and ubiquitin (Blesa et al. 2012). Effects from the loss of striatal dopamine and dysfunction of the nigrostriatal pathway resulted in a group of motor symptoms such as rigidity, tremor, bradykinesia and postural imbalance (Seet et al. 2010). Although the pathogenesis of this disease is still unclear, it is considered to be potentiated by the interaction between two factors; environment and genetic (Shafique et
al. 2011). It is well known that exposure to environmental herbicides is one of the environmental factors that have been reported to contribute to the pathogenesis of PD (Shafique et al. 2011). However, the risk of PD in human is reported to be higher in those with paraquat exposure and having the defective gene that encodes glutathione S-transferase T1 (GSTT1) (Goldman et al. 2012).

Paraquat (1,1 ‘dimethyl-4,4’-bipyridinium) is among the most commonly used herbicides in the agricultural sector worldwide (Cha et al. 2014). Paraquat (PQ) is a highly toxic quaternary ammonium herbicide that has been shown to produce toxicity particularly in the dopaminergic neurons of the rat and mouse brains (Blanco-Ayala et al. 2014; Yang & Tiffany-Castiglioni 2005). PQ is structurally similar to 1-methyl-4-phenylpyridinium ion (MPP+), a known neurotoxic and the active metabolite of 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine or MPTP (Brooks et al. 1999). Both PQ and MPTP ultimately cause the loss of dopaminergic neurons in the SN that leads to the clinical symptoms of Parkinsonism (Wesseling et al. 2001). PQ is considered as a potent oxidative stress inducer (Lascano et al. 2012). Oxidative stress plays a major role in the pathogenesis of Parkinson’s disease through the induction of nigral dopaminergic neuronal cells death (Nikolova et al. 2012). The death of these neuronal populations by PQ is mainly attributed to impaired mitochondrial functions as well as increased production of reactive oxygen species and reduction in oxygen consumption (Blanco-Ayala et al. 2014; Huang et al. 2016). PQ specific toxicity to dopaminergic neurons via oxidative stress mechanism in rats has prompted the current study to utilize the PQ-induced Parkinsonism model in order to study the antioxidant strategies of a herbal plant namely Zingiber zerumbet against Parkinsonism.

Herbal medicine has gained lots of attention due to the presence of various natural compounds that are beneficial to health including alleviating pathological symptoms. Zingiberaceae belong to order of Zingiberales, which forms an isolated group among the monocotyledons. Approximately 1000 species of Zingiberaceae are found in tropical Asia. In Peninsular Malaysia there are more than 160 species of Zingiberaceae identified (Abdul et al. 2008). Only a small portion of these ginger species have been cultivated for its use as vegetables, condiments, spices, flavors, dyes, medicines as well as ornaments (Habsah et al. 2000).

Zingiber zerumbet has been claimed to have high medicinal values (Yob et al. 2011). Zingiber zerumbet or locally known as lempoyang is used in local traditional medicine as a remedy for swelling, sores and loss of appetite (Abdul et al. 2008). The rhizome of Z. zerumbet has been used for anti-inflammatory, anti-ulceration, antioxidant, anti-pyretic, analgesic and anti-microbe (Somchit et al. 2005). This rhizome is reported to contain alkaloid, saponin, flavonoids and polyphenol as well as the volatile oil (Batubara 2013). The volatile oil of Z. zerumbet contains zerumbone as its main active constituent (Bhuiyan et al. 2009). Among the differences in the extraction methods employed, ethyl acetate extraction gives the highest yield of zerumbone (51.57%) (Budin et al. 2013). Zerumbone has versatile pharmaceutical properties including anti-inflammatory, antitumor, antioxidant, antimicrobial and antinociceptive activities (Somchit et al. 2012). Zerumbone has also been reported to inhibit proliferation of leukemic, cervical, ovarian, colon, breast and liver cancers (Abdelwahab et al. 2012, 2011).

As a neuroprotective agent, an administered substance of interest should be able to reverse some of the pathological damage or prevent further damage to the neuronal cells. A lot of study has found that active compounds present in the natural products have high contents of antioxidant and anti-inflammatory properties. These properties can protect the neurons against neurodegenerative conditions thereby preserving or even enhancing cognitive function (Essa et al. 2012; Hamid et al. 2018). Z. zerumbet extracts and the pure compound, Zerumbone, fit the ideal characteristics of a neuroprotective drug due to their anti-inflammatory, antioxidant and antiplatelet aggregate activities as reviewed in Yob et al. (2011).

In the current study, the main aim is to investigate the neuroprotective potential of ethyl acetate extract of Z. zerumbet rhizome through its antioxidants activities in paraquat-induced Parkinsonism in rats. The effects were determined by measuring the levels of oxidative stress markers (malondialdehyde (MDA), protein carbonylation), endogenous antioxidants (GSH, superoxide dismutase (SOD), catalase), histological changes of the neurons in the SN and behavioral changes of the neurons in the SN and behavioral analysis.

**MATERIALS AND METHODS**

**PLANT MATERIALS**

Fresh rhizomes of Z. zerumbet were collected from Temerloh, Pahang. The specimen was first validated and kept in the Herbarium, Faculty of Science and Technology (FST), Universiti Kebangsaan Malaysia (voucher number is UKMB-29952). The rhizomes were scrubbed and cleaned, chopped finely using dry blender and then air-dried at room temperature for three days.

**PLANT EXTRACTION**

The air-dried chopped rhizomes were soaked at room temperature in 100% n-hexane. After 72 h, the hexane containing the chopped rhizomes was filtered, the resulting filtrate (hexane) was kept for the next stage and the rhizome residue was soaked again for 72 h with a fresh hexane. This cycle was repeated 3 times. The rhizome residue from the third cycle of hexane was soaked with another solvent, 100% fresh ethyl acetate and later with 100% methanol, following the same exact procedure as hexane. The resultant solvent extracts (filtrates) collected at each stage were evaporated using rotavapor apparatus to yield dried crude extracts of hexane, ethyl acetate and methanol. All the crude extracts were stored at 4°C until further use.
Prior to use, the Z. zerumbet ethyl acetate extract was dissolved in 100% dimethyl sulfoxide (DMSO) and diluted in phosphate buffered saline (PBS; pH7.4), aliquoted and kept as stock at -20°C.

**EXPERIMENTAL PROTOCOL**

The procedures involving the use of laboratory animals were approved by the UKM Animal Ethical Committee (UKM/CEC) with the following approval code reference: FSK/BIOMED/2013/ASMAH/14-NOV.15566-NOV.-2013-JUNE-2014. Forty male Sprague-Dawley rats aged 6-8 weeks (140-200 g) were purchased from the UKM Laboratory Animal Resource Unit. The animals were housed in a controlled environment with room temperature at 24°C ± 1 and a 12-h light/dark cycle. Food and water were provided ad libitum. Rats were randomly divided into five groups with n=7/group. The groups were negative control group (normal saline), NAC (N-acetylcysteine) + PQ (20 mg/kg of NAC with PQ), PQ (PQ treated-group), 200 + PQ (200 mg/kg Z. zerumbet with PQ) and 400 + PQ (400 mg/kg of Z. zerumbet with PQ). Ethyl acetate extract of Z. zerumbet was given orally for 19 consecutive days and PQ was administered intraperitoneally (10 mg/kg) on Day 8-12 of the treatment regime. Both serum and fresh brains containing the SN region were taken for biochemical and histological analysis. Research timeline is shown in Figure 1.

**SAMPLE PREPARATION**

For the Z. zerumbet treatment group, the animals were killed 24 h after the last dose of Z. zerumbet extract on Day-19. Regardless of treatment, all animals were killed on Day-20. Prior to killing, the animals were anesthetized with KTX (Ketamine, xylazine and zoletil) with dosage of 0.5 mL/kg (Hamid et al. 2018) and decapitated to collect trunk blood and brains for sampling. Blood was collected in plain tubes and allowed to coagulate at room temperature for 3 h. Serum was obtained after centrifugation at 3000 rpm for 10 min and stored at -20°C until further use. The SN area was identified and isolated using brain matrices (Tedpella, USA) and washed with iced-cold KCl to wash off traces of blood. For histological purposes, the brain slice containing SN was fixed in 10% formalin, while for biochemical analysis; the SN was minced into small pieces, weighed and homogenized with 1:15% KCl solution at a ratio of 3 mL/g (v/w). The brain homogenate was then centrifuged at 4000 rpm for 20 min at 4°C. The supernatant obtained was then stored at -20°C until further analysis.

**BEHAVIORAL OBSERVATION**

**Tremor**  Tremor is one of the clinical hallmarks of PD. This behavioral study was performed to verify the Parkinsonism symptoms which were manifested in the PQ-induced rats. The phasic tremor of the whole body was evaluated and scored according to Coward et al. (1977); 0, no tremor; 1, occasional isolated twitches; 2, moderate or intermittent tremor associated with short periods of calmness; and 3, pronounced continuous tremor.

**BIOCHEMICAL ANALYSIS**

**Total Protein**  Total protein determination was based on Bradford (1976) method. Standard curve was prepared from a 500 µg/mL of bovine serum albumin (BSA) by pipetting 0.2, 0.4, 0.6, 0.8 and 1.0 mL of the solution into glass tube. The concentration of the measured proteins was plotted against the corresponding absorbance resulting in a standard curve which was used to determine the protein concentration in the samples. Depending on the concentration, diluted serum and SN supernatant were pipetted into test tubes separately. The volume was adjusted to 0.1 mL with PBS. A total of 5 mL of protein reagent was added to the test tubes and the contents were mixed by mild vortexing. The measurement of the absorbance was done at 595 nm against a reagent blank prepared from 0.1 mL of the appropriate buffer and 5 mL of protein reagent. The readings were taken between 2 min to 1 h.

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![Figure 1](image-url)  The experimental timeline. Zingiber zerumbet extract was given for 19 days consecutively via oral gavage. Then, on Day-8 until Day-12, the paraquat was injected daily (i.p) and the behavioural changes were observed. On Day-20, all the rats were sacrificed and sampling was done. Both fresh brains containing SN and serum were taken for further biochemical and histological analysis.
**Malondialdehyde (MDA) Level** The formation of thiobarbituric acid reactive substances (TBARS), which are the byproduct of lipid peroxidation, was measured according to the method by Ledwozyw et al. (1986). In this assay, the presence of reactive aldehyde such as MDA (end product of lipid peroxidation) reacts with thiobarbituric acid (TBA) reagent under an acidic condition to generate a pink coloured product. A total of 0.1 mL of SN homogenate supernatant was added to 0.4 mL of distilled water. The mixture was then added with 2.5 mL of trichloroacetic acid (TCA) and left to incubate at room temperature for 15 min. Then 1.5 mL of TBA was added and the resultant mixture was heated in a water bath at 100°C for 30 min or until a faint pink coloured solution was obtained. After cooling, the coloured product was added with 1 mL of butanol, vortexed for 3 min and the top layer of the resulting mixture was taken and the absorbance was measured using spectrophotometer at 532 nm.

**Protein Oxidation Level (Protein Carbonyl, (PC))** Protein oxidation was evaluated by the determination of protein carbonyl level. Dinitrophenhydrazine (DPNH) reacts with protein carbonyl groups to produce the stable 2,4-dinitrophenyl (DNP) hydrazone protein adducts that can be detected spectrophotometrically at 370 nm according to the method by Levine et al. (1990). In short, proteins were precipitated by the addition of 20% TCA and were dissolved in (how much of DPNH). The results were reported as nmol/mg protein.

**Superoxide Dismutase (SOD) Activity** Superoxide dismutase activity was measured according to the method of Beyer and Fridovich (1987). In summary, the reaction mixture consisting of 50 mM potassium phosphate buffer pH7.8, 0.1 mM EDTA, 9.9 mM L-methionine, 5.7 x 10^-3 % nitro blue tetrazolium (NBT) and 2.5 x 10^-3 % (w/v) Triton X-100 and riboflavin (0.01 mL of 4.4%) were freshly prepared on the day of the assaying. A total of 1.0 mL of the reaction mixture was added to 20 μL of SN homogenate in a cuvette to initiate the reaction which was then placed in a box illuminated with 20 W neon lamps for 7 min. The reduction of NBT was measured at 560 nm spectrophotometrically. The results were calculated by subtracting the changes in absorbance of the blank from the sample and dividing ΔA by 0.012 to obtain the McCord-Fridovich units of activity.

**Catalase (CAT) Activity** Catalase activity was determined as described by Aebi (1984). A total of 50 mM phosphate buffer (pH7.0) and 30% H₂O₂ were freshly prepared on the day of the assaying. Two mL of sample solution was mixed with 1 mL of H₂O₂. The decomposition of H₂O₂ was measured at 240 nm via spectrophotometer using a blank containing 2 mL of the sample solution and 1 mL of phosphate buffer. The reaction time was ensured to be within 30 s.

**Glutathione Peroxidase (GPx) Activity** Glutathione peroxidase measurement was determined at 340 nm spectrophotometrically using the method of Paglia and Valentine (1967). Both the blank and the system cuvettes contained 0.1 M KPO₄ buffer (pH7.0), 2 μM EDTA, 10 units/mL glutathione reductase, 4 mM sodium azide, 200 mM NADPH and sample. In addition, the system cuvette contained 1.0 mM glutathione, GSH. After 10 min of pre-incubation at 37°C, the reaction was started by adding 1.0 mM H₂O₂ to the blank and system cuvettes. In order to correct the non-enzymatic oxidation of GSH and NADPH by H₂O₂, an additional blank assay in which the buffer was substituted for the sample was performed.

**Glutathione (GSH) Level** The assay of GSH with DTNB was performed by following Ellman’s method (1959). Equal volume of 20% trichloroacetic acid (TCA) containing 1 mM EDTA was added to the brain homogenate to precipitate the tissue protein. After 5 min, the resulting mixture was centrifuged for 10 min at 200 rpm. A total of 200 μL of the supernatant was transferred to a new set of test tubes and added with 1.8 mL of the Ellman’s reagent, 5, 5’-dithio bis2nitrobenzoic acid (DTNB). The resulting solution was measured at 415 nm against a blank. Absorbance values were compared with a standard curve generated. The GSH level was measured in nmol/mg protein.

**HISTOLOGICAL ANALYSIS**

Histological examination on the region of interest of the brain was performed according to routine histological techniques. Briefly, after the animal was killed, the brain containing the SN region was isolated and harvested, rinsed in normal saline and sectioned into small pieces. The sectioned tissues were then fixed in 10% formalin, dehydrated in stepwise increasing concentration of ethanol solution (50% to 100%) and later embedded in paraffin. Tissue sections of 5 μm thickness were produced using microtome and fixed overnight on slides. The slides were subsequently stained with hematoxylin and eosin (H&E) and then analyzed under a light microscope (Motic Images, Hong Kong).

**STATISTICAL ANALYSIS**

Statistical analysis was done using SPSS version 20.0 (IBM Corp., AS). Data was first analyzed using Shapiro-Wilk normality test. Later, one way analysis of variance (ANOVA) was employed to compare means between groups followed by posthoc Tukey test. Mixed ANOVA was used to analyze the differences in the tremor score. Data was expressed as means ± standard error of mean (SEM) with p<0.05 was considered as statistical significance.

**RESULTS**

**TREMOR OBSERVATION**

Tremors observed in the rats were evaluated and scored. In short, a score of 0 means absence of tremor, 1 indicates
occasional isolated twitches being observed, 2 indicates moderate or intermittent tremor associated with short periods of calmness, whereas a score of 3 indicates that the rats showed pronounced continuous tremor. Higher score indicates the severity of the symptoms shown (Figure 2). There was a noted decline in the tremor score for both 200 and 400 mg/kg extract group compared to the PQ-treated group as seen in the post-induction period. Administration of Z. zerumbet in both the extract groups alleviated the severity of the tremors caused by PQ induction.

EFFECTS OF Z. ZERUMBET EXTRACT ON OXIDATIVE STRESS MARKERS

Administration of PQ for five consecutive days induced oxidative stress in the SN area as indicated by a significant increase in the level of protein carbonyl (Figure 3(B)). Induction of PQ in the Z. zerumbet treatment groups of both 200 + PQ and 400 + PQ showed significantly reduced levels of MDA and protein carbonyl when compared to the PQ-treated group (p<0.05).

EFFECTS OF Z. ZERUMBET EXTRACT ON ANTIOXIDANT STATUS

Administrations of PQ for five consecutive days also caused a significant reduction in the activities of CAT (Figure 3(D)) and GPx (Figure 3(E)) in the SN. In contrast, administration with either 200 or 400 mg/kg of Z. zerumbet extract significantly prevented the decreased in the enzyme levels and improved the activities of SOD (Figure 3(C)), CAT (Figure 3(D)) and GPx (Figure 3(E)) (p<0.05). In the serum, paraquat induction caused a significant reduction of SOD (Figure 4(A)) and CAT (Figure 4(B)) enzyme activities compared to the normal control group (p<0.05). However, administration of Z. zerumbet at 400 mg/kg increased both of these enzyme activities significantly when compared to the PQ-treated group (p<0.05).

HISTOLOGICAL OBSERVATION

Histological sections of the brain sample containing the SN region was scrutinized under a light microscope. There was presence of fibrillary aggregates called Lewy bodies (LBS) in the SN of the PQ-treated group (Figure 4(D)). In our observation, administration of PQ caused the formation of fragmentation in the nucleus of the neurons and the presence of Lewy bodies. In contrast, administration of Z. zerumbet extract prevented the nucleus from undergoing fragmentation and thus appeared intact and normal (Figure 4(E)).

DISCUSSION

In PD, there is growing evidence that oxidative stress mechanism play a role in causing neuronal cell death. In this study, administration of 10 mg/kg of PQ for five consecutive days was able to induce Parkinsonism in rats as demonstrated by behavioral observation (tremor). Co-administration of ethyl acetate extract of Z. zerumbet with PQ showed that there was a reduction in the oxidative stress level in Parkinsonism rats compared to the group that was given only PQ. Furthermore, the protective effect of Z. zerumbet was found to be more remarkable in the 400 + PQ group than in the 200 + PQ group. This finding is in line with previous study conducted by Hamid et al. (2011) on hepatoprotective effects of the same extract. In that study, they have shown that 400 mg/kg of Z. zerumbet ethyl acetate extract gave a better hepatoprotective effects to the paracetamol-induced hepatotoxicity rats compared to the 200 mg/kg dosage.

Oxidative stress is an imbalance between the production of ROS and an antioxidant defense system to detoxify the reactive intermediates or to repair the resulting damage (Betteridge 2000). Excessive in ROS production or impaired antioxidant system will lead to oxidative

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![Figure 2](image-url)

**Figure 2.** Tremor score. Tremor is evaluated based on scoring, the higher the score, the more severe the symptoms shown. All animals scored 0 prior to paraquat induction. Based on the graph, there was a decreased in the tremor score for both the extract supplemented groups compared to PQ-treated group. Administrations of Z. zerumbet from Day 1 - Day 19 (200 mg/kg and 400 mg/kg) alleviated the severity of the tremor during the induction period. Mixed ANOVA test revealed significant differences in all groups (p<0.05) during paraquat induction (Day 8-Day 12).
imbalance and can cause ROS overproduction. Disruption in the normal redox state of cells can cause toxic effects through the production of peroxides and free radicals that damage all components of the cell, including proteins, lipids and DNA (Poljsak et al. 2013).

In this study, both MDA and the PC levels were observed to increase in the PQ-treated group, with only the PC level was statistically proven to be significantly higher compared to the normal control group. Malondialdehyde (MDA) is one of the most frequently used indicator of lipid
peroxidation due to its stability. Lipid peroxidation is one of the major products of free radical-mediated injury to brain because of high concentration of polyunsaturated fatty acid in the brain compared to the other organs (Montine et al. 2002). Malfunction of the antioxidant defense system causes excessive lipid peroxidation process that eventually elevates the level of MDA. The increment in the MDA level is considered as an indicator of cellular damage (Kaplowitz 2000).

Study by Giasson et al. (2002) reported that the marker for oxidative stress such as lipid peroxidation products was increased in the brains of PD patients. Moreover, according to Perfeito et al. (2012), analysis of biochemical markers of oxidative damage in brain tissue samples and postmortem PD patients showed increased levels of MDA and 4-hydroxynonenal in the SN area further confirmed the occurrence of lipid peroxidation. Due to highly electrophilic properties that can react with other biomolecules such as protein and DNA, MDA is considered as a highly toxic molecule that can damage other molecules and eventually causes cell death (Thanan et al 2015). The presence of carbonyl group in proteins may indicate that the proteins are exposed to oxidative stress damage. Alzheimer’s disease, chronic lung disease, chronic renal failure, diabetes and sepsis are examples of disorders which are associated with an increase in protein carbonyl content in the tissue (Dalle-Donne et al. 2003; Kadiiska et al. 2005).

In this study, we have proven that administration of ethyl acetate extract of *Z. zerumbet* at both 200 and 400 mg/kg can reduce the level of MDA and PC compared with the PQ-treated group. The ability of these extracts to protect the neurons against lipid peroxidation is supported by studies from Ruslay et al. (2007). The findings of the study reported that ethyl acetate extract of *Z. zerumbet* able to inhibit lipid peroxidation through its radical scavenging activities. Previous studies on hepatoprotective (Hamid et al. 2011) and nephroprotective (Abdul Hamid et al. 2012) also corroborated the findings as both studies showed that *Z. zerumbet* ethyl acetate extract reduced the
level of MDA in both the liver and kidney of chemically-induced rats.

In order to protect our body from the free radical toxicity, the production of free radicals are controlled by cellular antioxidant defense mechanisms which include base-enzymatic and non-enzymatic antioxidants (Nordberg & Arner 2001). Antioxidants are substances that can prevent or slow the oxidation process. The function of antioxidant is to neutralize excess free radicals, protecting the cells against the toxic effects and for disease prevention (Sailaja Rao et al. 2011). Antioxidant enzymes are the first defense systems of cells against oxidative damage.

Activities of CAT and GPx in the SN of the PQ-treated group were significantly lower than the normal control group (negative control). The main key players that provide protection against oxidative stress in the brain are SOD, CAT and GPx (Nikolova 2012). Toxins formed in the brain are usually removed or deactivated by a variety of mechanism of protection. This mechanism however, is compromised in Parkinson’s disease when there is a rapid drop in the activities of CAT, GPx and GSH concentrations in the SN (Ogunro et al. 2014). With the failure of the protective mechanism, oxidative stress would easily occur and further causes tissue damage (Chinta et al. 2007). In addition, the activities of SOD and CAT enzymes in the rat serum of PQ-treated group were also lower compared to the normal control group. These results were consistent with the study conducted by Sanyal et al. (2011), in which PD patients have lower antioxidant activity as indicated by significantly decreased level of SOD, CAT and GPx suggesting that the reduction of these enzymes have a correlation with the occurrence of the disease.

There were increased in the activities of SOD, CAT and GPx enzymes in the group of rats given ethyl acetate extract of Z. zerumbet. The increment proves that there is an overall enhancement of the endogenous antioxidant defense system in the PQ-treated rats. The results obtained are in line with study by Hamid et al. (2011). In their study, treatment with Z. zerumbet at 200 and 400 mg/kg were also able to increase the superoxide dismutase (SOD) activities in the liver homogenate. The antioxidant activities might be attributed to zerumbone, the main bioactive compound found in the ethyl acetate extract of Z. zerumbet. Zerumbone, a sesquiterpene compound and also a potent antioxidant, may indirectly induce glutathione biosynthesis and therefore provides intracellular protection mechanisms through free radical scavenging activity (Ibrahim et al. 2010; Nakamura et al. 2004). However, the increment was more significantly favorable to the 400 mg/kg group compared to 200 mg/kg. This may be due to excessive production of free radicals following PQ exposure in the SN and the effect of 200 mg/kg extract of Z. zerumbet was insufficient to overcome the detrimental effects of the generated ROS. In all, Z. zerumbet proves to have high antioxidant activity through induction of endogenous antioxidant that reduces free radical activity and this can be evidenced by the results of this study. Similarly, this finding is also supported by Hamid et al. (2018) that proved the ability of ethyl acetate extract of Z. zerumbet in protecting ethanol-induced brain damages via its antioxidant property.

Substantia nigra (SN) which is located in the midbrain plays a role in controlling motor function. The leading cause of PD is the loss of dopaminergic neuron in substantia nigra pars compacta (Perfeito et al. 2012). Dopaminergic neurons are important to ensure normal motor function. Based on histological observations of the SN, there was a karyolytic feature in the nuclei of neurons in the PQ-induced tissues. Karyolysis is one the characteristics of cell death. According to Gujral et al. (2002), 99% of cell death is characterized by cell lysis, vacuole formation, karyolysis and karryorhexis (Gujral et al. 2002). Cell death can be due to apoptosis or necrosis. This is supported by research done by Somayajulu-Nitu (2009), which showed that PQ at the dose of 10 mg/kg/week for 3 weeks had resulted in the death of 65% dopaminergic neurons in the SN. Similarly, a study conducted by Shimizu et al. (2003) showed that subchronic administration of PQ at the dose of 10 mg/kg for 5 days was capable of causing death to dopaminergic neurons.

Histological features for ethyl acetate extract of Z. zerumbet at the dose of 400 mg/kg showed that there was no fragmentation detected in the nucleus of the neurons. This indicates that ethyl acetate extract of Z. zerumbet at this particular dose may prevent neuronal cell death from occurring. Besides that, the histological hallmark of Parkinson's disease (PD) is the presence of fibrillar aggregates called Lewy bodies (LBS). Formation of LBS is considered to be the indicator for neuronal degeneration because the neuronal loss is found in the predilection sites for LBS (Wakabayashi et al. 2007).

**Conclusion**

Based on the results obtained, it can be concluded that administration of the ethyl acetate extract of Z. zerumbet provides neuroprotective effect on PQ-induced Parkinsonism in rats. The inhibition of oxidative stress was evidenced by reduction of lipid and proteins oxidation and enhanced antioxidant enzyme activities (SOD, CAT and GPx). The most significant effect was observed with the administration of 400 mg/kg of extract compared to a lower dose of 200 mg/kg. This finding indicates the difference in the presence and amount of antioxidant compounds that contribute significantly to Zingiber zerumbet antioxidant property.

Ethyl acetate extract of Z. zerumbet at the dosage of 200 mg/kg and 400 mg/kg have been found to prevent oxidative stress and neuronal damage induced by paraquat and therefore, can be developed as a preventive and therapeutic agent for neurodegenerative-induced environmental toxins. The authors have declared no conflict of interest.
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