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(+)-Neocadambine A and (-)-Nauclederine Isolated from the Bark of *Neolamarckia* cadamba (Rubiaceae) as Natural Advanced Glycation End Products (AGEs) Inhibitors

((+)-Neocadambine A dan (-)-Nauclederine Diasingkan daripada Kulit *Neolamarckia cadamba* (Rubiaceae) sebagai Perencat Produk Akhir Glisasi Lanjutan Semula Jadi (AGEs))

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ABSTRACT

The phytochemical study on the dichloromethane extract of *Neolamarckia cadamba* (Roxb.) Bosser has afforded two indole alkaloids, (+)-neocadambine A (1) and (-)-nauclederine (2). Their structures were confirmed by extensive spectroscopic analysis and by comparing with the reported data. (+)-Neocadambine A (1) and (-)-nauclederine (2) exhibited potent inhibition activity of advanced glycation end products (AGEs) with IC₅₀ values of 1.2 and 0.95 mM, respectively, while the latter was almost two times more potent than the standard, aminoguanidine (1.8 mM). This is the first report on the compounds isolated from this plant with AGEs inhibition activity. In addition, (-)-nauclederine (2) was isolated for the first time in the genus of Neolamarckia. Complete ¹H-NMR and ¹³C-NMR of compound **2** were also reported.

Keywords: Advanced glycation end products; indole alkaloid; (-)-nauclederine; (+)-neocadambine A; *Neolamarckia cadamba*

ABSTRAK

Kajian fitokimia ke atas ekstrak dikloromethana bagi *Neolamarckia cadamba* (Roxb.) Bosser telah menghasilkan dua alkaloid indola, (+)-neocadambine A (1) dan (-)-nauclederine (2). Strukturnya dikenal pasti melalui pelbagai analisis spektroskopi dan perbandingan dengan data yang telah dilaporkan. (+)-Neocadambine A (1) dan (-)-nauclederine (2) menunjukkan aktiviti perencatan kuat terhadap produk akhir glisasi lanjutan (AGE) dengan nilai IC₅₀ masing-masing 1.2 dan 0.95 mM sementara (-)-nauclederine (2) hampir dua kali ganda lebih kuat daripada piawai, aminoguanidine (1.8 mM). Ini adalah laporan pertama mengenai sebatian yang diasingkan daripada tumbuhan ini dengan aktiviti perencatan terhadap AGE. Selain itu, (-)-nauclederine (2) diasingkan untuk pertama kalinya dalam genus Neolamarckia. ¹H-NMR dan ¹³C-NMR yang lengkap bagi sebatian **2** juga dilaporkan.

Kata kunci: Alkaloid indola; (-)-nauclederine; (+)-neocadambine A; *Neolamarckia cadamba*; produk akhir glisasi lanjutan

Introduction

Rubiaceae is one of the largest families of angiosperms and consists of approximately 600 genera and more than

10,000 species (Silva et al. 2010). The Rubiaceae family is characterized by the production of bioactive metabolites such as alkaloid as secondary metabolite with diverse

pharmacological potential (Martins & Nunez 2015). Examples of pharmacological properties exhibited by the metabolites of Rubiaceae plants are antimicrobial (Sultana et al. 2015), anti-diabetic (Habtemariam 2019) and anti-inflammatory activities (Qureshi et al. 2011).

Neolamarckia cadamba (Roxb.) Bosser (Rubiaceae), locally known as Laran, is a tree of moderate size found in Malaysia (Qureshi et al. 2011). N. cadamba is widely used in Indian traditional formulations in which the leaves decoction was consumed for the treatment of ulcers and wounds since many years ago (Verma et al. 2018). According to Pandey and Negi (2016), the fruit has a cooling effect and has been used for quenching thirst during high fever as prescribed by Charaka Samhita (Pandey & Negi 2016). Besides, the fruit of N. cadamba has also been used in different food preparations by tribal people (Pandey & Negi 2016).

Various extracts of *N. cadamba* such as ethanol and hydroethanolic extracts have beneficial effects in reducing the elevated blood glucose level of hyperglycemic mice (Ahmed et al. 2011; Alam et al. 2011). For example, an anti-diabetic study showed that the hydroethanolic fruit extract (400 mg/kg) had higher hypoglycemic activity than standard drug glibenclamide (0.6 mg/kg) in alloxan induced diabetic Swiss albino rats (Alam et al. 2011).

Maillard reaction which often known as nonenzymatic browning reaction is implicated in the development of pathophysiology in age-related diseases such as diabetes mellitus (Thorpe & Baynes 1996). Maillard reaction led to advanced glycation end products (AGEs) through a series of non-enzymatic, sequential and parallel reactions, which can be divided into two stages. The early glycation was reversible and involves the production of a Schiff base from the carbonyl group of a reducing sugar and the primary amino groups of a protein (lysine, arginine). The imine adduct undergoes rearrangement to form Amadori products such as HbA1c (glycated haemoglobin), which widely used as a diabetes control marker (Wolffenbuttel et al. 1996). During the late stage, complex irreversible oxidation, dehydration and cyclization reactions lead to AGEs via intra- and intermolecular protein cross-linking (Boisard et al. 2014; Peyroux & Sternberg 2006; Poulsen et al. 2013; Reddy & Beyaz 2006).

AGEs accumulation occurs in healthy person but the AGEs level is higher for those patients who have hyperglycemia and diabetes or in condition involving oxidative stress (Luevano-Contreras & Chapman-Novakofski 2010; Vlassara & Uribarri 2014). AGEs are associated with many pathogenic disorders such as pathogenesis of diabetes (Brownlee 2001; Singh et al. 2001; Wada & Yagihashi 2005), atherosclerosis (de Leeuw et al. 2005; Jandeleit-Dahm & Cooper 2008), neurological diseases such as Alzheimer's disease (Takeuchi & Yamagishi 2008) or joint diseases (DeGroot 2004). AGEs are also responsible for aging, tissue and skin damages (Grillo & Colombatto 2008) as well as autoimmune diseases (Kurien et al. 2006). This consideration has driven the scientific community to identify and develop new AGE inhibitors that are able to prevent oxidation using free radical scavengers or transition metal chelators in order to trap reactive dicarbonyl species or to break AGE cross-linking (Elosta et al. 2012; Reddy & Beyaz 2006). By addressing to this issue, finding natural products possess anti-AGE properties with free of or fewer side effects are critically needed.

Our group has conducted an AGEs inhibitory assay on several plant extracts and the dichloromethane extract of the bark of *N. cadamba* displayed a strong anti-AGEs effect similar to the standard, aminoguanidine (0.15 mg/mL). This result has prompted us to conduct the present study. Hence, we shall report the isolation and elucidation of (+)-neocadambine A and (-)-nauclederine (2) together with their anti-AGEs activity. To the best of our knowledge, this is the first report on the anti-AGEs effect of indole alkaloid extracted from *N. cadamba*.

MATERIALS AND METHODS

GENERAL EXPERIMENTAL PROCEDURES

The 1D-and 2D-NMR spectra were recorded deuterated methanol (CD,OD) and deuterated pyridine (C,D,N) BRUKER Advance III 600 NMR spectrometers. Chemical shifts $(\delta_{\Pi}$ and $\delta_{C})$ are expressed in ppm and J values in Hz. The HR-ESI-MS spectra were obtained from Agilent 6530 Accurate-Mass Q-TOF with electrospray ionization (ESI) (Santa Clara, CA, USA). Ultraviolet spectra were obtained in spectroscopic grade methanol with Shimadzu UV-250 UV-visible spectrometer. IR spectra were recorded on a Perkin Elmer Spectrum 1600 FTIR Spectrometer with spectroscopic grade chloroform as the solvent. A Jasco P-1020 polarimeter was used to record the optical rotation. Recycling High Performance Liquid Chromatography (RHPLC) with JAIGEL-ODS-AP-30, SP-120-15 reversed phase column was used for purification of compound. In the column chromatography, silica gel 60 F_{254} was used for thin layer chromatography (TLC). Silica gel 60 F₂₅₄ plates backed by glass and aluminium were used for TLC. All solvents are AR grade except those used for bulk extraction.

PLANT MATERIAL

The bark of *N. cadamba* (Roxb.) Bosser was collected from Empangan Sg. Sunda Jeli, Kelantan, Malaysia. The voucher specimens (KL 5696) were deposited at the Herbarium of Department of Chemistry, Faculty of Science, Universiti Malaya, Kuala Lumpur, Malaysia.

EXTRACTION, FRACTIONATION, PURIFICATION AND ISOLATION OF COMPOUNDS

Dried and ground bark of the N. cadamba (1.8 kg) were defatted with hexane (10 L) for three days at room temperature. The hexane extract was filtered and dried using rotary evaporator. After that, the plant materials were moistened with ammonia solution and soaked for 4 h. The plant material was then re-extracted with dichloromethane (30 L) for three days. After the extract was filtered and dried using rotary evaporator, 17.5 g of dichloromethane extract were obtained. The plant materials were soaked again with methanol (28 L) for three days and the extract was dried using rotary evaporator. An amount of 70.0 g of methanol extract was obtained. The dichloromethane extract (1.75 g) was subjected to column chromatography over silica gel using dichloromethane and methanol solvent (100:0, 95:5, 90:10, 80:10 and 0:100) to obtain 5 fractions. Further purification of fraction 3 (20 mg/mL) using RHPLC yielded alkaloid 1 (4.9 mg, flow rate 5 mL/min: isocratic: 100% MeOH, injection volume 3 mL). Alkaloid 2 (7.7 mg, MeOH-CHCl₂; 100:0) was purified from fraction 3 by column chromatography C18.

(+)-Neocadambine A (1): yellow amorphous solid; +17.14° (c 0.07, MeOH); UV (MeOH) λ_{max} 214, 244, 291, 313 nm; ¹H NMR and ¹³C NMR see Table 1; HRESIMS m/z 366.1466 [M + H]⁺ (calcd for $C_{20}H_{20}N_3O_4$, 366.1454).

(-)-Nauclederine (2): yellow amorphous solid; -6.67° (c 0.06, MeOH); UV (MeOH) $\lambda_{\rm max}$ 213, 227, 305 nm; ¹H NMR and ¹³C NMR see Table 2; HRESIMS m/z 322.1546 [M + H]⁺ (calcd for $C_{19}H_{20}N_3O_2$, 322.1556).

ADVANCED GLYCATION END PRODUCTS (AGES) INHIBITORY ACTIVITY

IC₅₀ values were determined as described by Derbré et al. (2010) and Séro et al. (2013). Briefly, bovin serum albumin (BSA) (10 mg/mL) was incubated with D-ribose (0.5 M) together with the tested compound

(3 μM to 3 mM) or extract (1 μg to 1 mg) in 50 mM phosphate buffer at pH 7.4 (NaN₃, 0.02%). Solutions were incubated in 96-well microtiter plates at 37 °C for 24 h in a closed system before AGE fluorescence measurement. Fluorescence resulting from the incubation, under the same BSA (10 mg/mL) and tested compound (3 μM to 3 mM) or extract (1 μg to 1 mg) conditions, was subtracted for each measurement. A control (i.e. no inhibition of AGE formation) consists of wells with BSA (10 mg/mL) and D-ribose (0.5 M). A blank control (i.e. 100% inhibition of AGE formation) consists in wells with only BSA in buffer (10 mg/mL). The final assay volume was 100 μ L. Pentosidine-like (λ_{exc} 335 nm, λ_{em} 385nm) AGE fluorescence was measured using a microplate spectrofluorometer. Aminoguanidine was used as a standard. In this type of automation, a single analysis is sufficient for an accurate IC₅₀ determination (Derbré et al. 2010; Séro et al. 2013). The percentage of AGE formation for each compound or extract concentration was calculated as follows: Dose-effect curves are best fitted with a sigmoidal dose response equation using Sigma Plot 12.0 software which enables calculation of the IC₅₀ values.

RESULTS AND DISCUSSION

Compound 1 was isolated as a yellowish amorphous solid with the molecular formula of C₂₀H₁₉N₃O₄, as determined by molecular ion peak positive-mode HR-ESI-MS which showed a pseudomolecular ion peak $[M+H]^+$ at m/z 366.1466 (calcd. 366.1454). The UV spectrum showed absorption bands at 313, 291, 244, and 214 nm. The IR spectrum exhibited absorption at 1642 cm⁻¹ indicating the presence of lactam carbonyl functionality (Hu et al. 2009). Thorough analysis of all spectra data (Table 1) and comparison with literature data (Yuan et al. 2020) led to the conclusion that compound 1 has the similar structure with neocadambine A (Figure 1). However, the optical rotation signs of compound 1 is positive which is opposite to the sign in neocadambine A. This difference implied that compound 1 is the enantiomer of neocadambine A. Hence, the relative configuration of C-19 in compound 1 will be the opposite of that in neocadambine A. The connectivity between all the carbons and protons were established thorough analysis of COSY, HSQC and HMBC spectra (Table 1, Figure 2). H-19 in compound 1 appeared as doublet of doublet with coupling constant of 7.0 and 4.0 Hz, respectively. Besides, neocadambine A was reported

with negative optical rotation while compound 1 is with positive optical rotation. Hence, compound 1 is named as (+)-neocadambine.

Compound 2 was isolated as a yellowish amorphous solid. The molecular formula of $C_{19}H_{10}N_3O_2$ was

determined by positive HR-ESI-MS which showed a pseudomolecular ion peak [M+H]⁺ at m/z 322.1546 (calculated 322.1556). The UV spectrum showed absorption at 305, 227, and 213 nm. In the IR spectrum, an absorption band of the NH stretching vibration was

FIGURE 1. Structures of (+)-neocadambine A (1) and (-)-nauclederine (2)

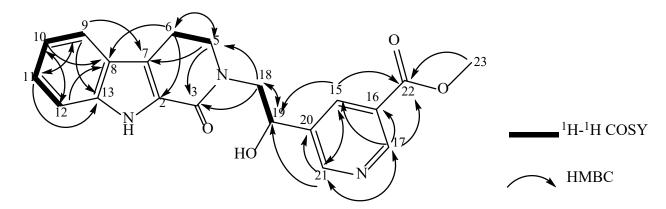


FIGURE 2. ¹H-¹H COSY and HMBC correlations of 1

observed at 3250 cm⁻¹. The ¹H-NMR spectrum (Table 2) of this alkaloid showed the presence of indole moiety by exhibiting two doublets signals at $\delta_{\rm H}$ 7.47 (1H, d, J = 7.7 Hz, H-10) and $\delta_{\rm H}$ 7.20 (1H, d, J = 7.7 Hz, H-13) as well as two triplet of doublets signals at $\delta_{\rm H}$ 7.00 (1H, td, J = 7.7, 1.1 Hz, H-11) and $\delta_{\rm H}$ 7.04 (1H, td, J = 7.7, 1.1 Hz, H-12) in the aromatic region (Wang et al. 2015). Besides, three proton signals were observed in the deshielded region at $\delta_{\rm H}$ 8.16 (1H, t, J = 2.0 Hz, H-16), $\delta_{\rm H}$ 8.96 (1H, d, J = 2.0 Hz, H-18) and $\delta_{\rm H}$ 8.55 (1H, d, J = 2.0 Hz, H-20). These three proton signals; H-16, H-18 and H-20 correlated with the carbon signals of $\delta_{\rm C}$ 138.9, $\delta_{\rm C}$ 149.5 and $\delta_{\rm C}$ 154.3, respectively, in the HSQC spectrum, thus implying the presence of 3,5-disubstituted pyridine

moiety. In addition, a singlet proton signal was observed at $\delta_{\rm H}$ 3.90, corresponding to the methoxy group of C-22 ($\delta_{\rm C}$ 53.1). The 13 C-NMR (Table 2) and DEPT spectra showed nineteen carbon signals, i.e. one carbonyl carbon, six quaternary carbons, eight methines, three methylenes and one methyl. The HMBC spectrum (Figure 3) showed correlations of H-4 with C-2 and C-6, H-6 with C-4 and C-8, H-7 with C-2 and C-8 indicated that the azepane unit is fused to the indole ring through C-2 and C-8. Moreover, the methoxy proton, H₃-22 correlated with the carbonyl carbon of C-21 ($\delta_{\rm C}$ 167.0) which can be observed in HMBC spectrum signifying the existence of methyl ester group. The linkage of this methyl ester group to the pyridine (ring D) through C-17 ($\delta_{\rm C}$ 128.0) was

supported by the HMBC correlations of H-18 with C-17, H-16 and H-18 with C-21. Furthermore, correlations of H-4 with C-3 and C-15, H-3 with C-16 and C-20, H-20 with C-15, H-16 and H-20 with C-3 indicated that the pyridine unit was connected to the azepinoindole skeleton through C-3. From the analysis of the spectroscopic data

obtained and by comparison with the limited reported data, the structure of (-)-nauclederine (2) was established. This compound was previously isolated from *Nauclea diderrichii* in 1971 and had been synthesized in 1976 (McLean et al. 1976; Murray et al. 1972). However, there is no thorough spectroscopic data being reported.

TABLE 1. ¹H-NMR (600 MHz) and ¹³C-NMR (150 MHz) of (+)-neocadambine A (1) in C₅D₅N

Position	$^{1}{ m H}$ $ \delta_{ m H}$ (multiplicity, J in Hz)	13 C ($\delta_{\rm c}$)		$\mathrm{HMBC}(\mathrm{H}{\rightarrow}\mathrm{C})$
		In ppm	Туре	
NH-1	14.51 (s)	-	-	
2	-	128.2	Quaternary	
3	-	170.6	Quaternary	
5a	4.09-4.12 (m)	54.2	Methylene	C3, C6, C7
5b	4.20-4.23 (m)			
6	3.10-3.16 (<i>m</i>)	20.2	Methylene	C2, C5, C8
7	-	124.5	Quaternary	
8	-	124.7	Quaternary	
9	7.55 (<i>d</i> , 7.9)	122.3	Methine	C7, C11, C13
10	7.14 (<i>t</i> , 7.9)	122.3	Methine	C8, C12
11	7.42 (<i>t</i> , 7.9)	129.6	Methine	C9, C13
12	8.19 (<i>d</i> , 7.9)	115.0	Methine	C8, C10
13	-	143.2	Quaternary	
15	8.94 (<i>t</i> , 1.9)	136.1	Methine	C19, C21, C22
16	-	126.6	Quaternary	
17	9.39 (<i>d</i> , 1.9)	150.9	Methine	C15, C16, C21, C22
18a	4.89 (dd, 13.8, 4.0)	62.3	Methylene	С3
18b	5.10-5.30 (overlapping signal)			C3, C5, C19
19	5.96 (dd, 7.0, 4.0)	69.9	Methine	C18
20	-	137.9	Quaternary	
21	9.59 (<i>d</i> , 1.9)	153.4	Methine	C15, C17, C19, C20
22	-	166.2	Quaternary	
23-OCH ₃	3.83 (s)	52.6	Methyl	C22

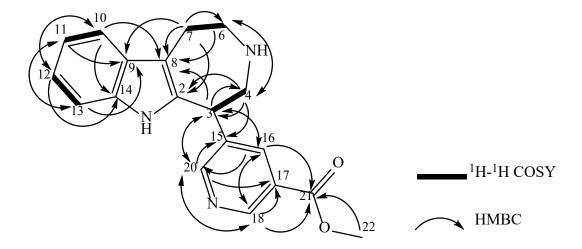


FIGURE 3. ¹H-¹H COSY and HMBC correlations of 2

TABLE 2. $^{1}\text{H-NMR}$ (600 MHz) and $^{13}\text{C-NMR}$ (150 MHz) of (-)-nauclederine (2) in CD_{3}OD

Position	$^{1} ext{H}$ $\delta_{_{ ext{H}}}$ (multiplicity, J in Hz)	$^{13}\text{C} \left(\delta_{\text{c}}\right)$		$HMBC(H{\rightarrow}C)$
		In ppm	Туре	
2	-	136.4	Quaternary	
3	4.50 (dd, 4.9, 3.4)	46.5	Methine	C4, C8, C16, C20
4a	3.36 (dd, 13.7, 3.3)	56.2	Methylene	C2, C6, C15
4b	3.45 (<i>dd</i> , 13.7, 4,9)			C2, C3, C6, C15
6a	2.98-3.02 (m)	51.1	Methylene	C4, C8
6b	3.27-3.30 (m)			C4, C8
7	3.05-3.09(m)	28.0	Methylene	C2, C6, C8, C9
8	-	114.4	Quaternary	
9	-	130.2	Quaternary	
10	7.47(d, 7.7)	119.0	Methine	C8, C12, C14
11	7.00 (td, 7.7, 1.1)	119.9	Methine	C9, C13
12	7.04 (td, 7.7, 1.1)	122.5	Methine	C10, C14
13	7.20 (d, 7.7)	111.8	Methine	C9, C11
14	-	137.3	Quaternary	
15	-	140.0	Quaternary	
16	8.16 (<i>t</i> , 2.0)	138.9	Methine	C3, C18, C20, C21
17	-	128.0	Quaternary	
18	8.96 (<i>d</i> , 2.0)	149.4	Methine	C16, C17, C20, C21
20	8.55 (<i>d</i> , 2.0)	154.3	Methine	C3, C15, C17, C18
21	-	167.0	Quaternary	
22	3.90 (s)	53.1	Methyl	C21

Both compounds; (+)-neocadambine A (1) and (-)-nauclederine (2), have been tested for AGEs inhibitory effect and the IC_{50} values calculated are 1.2 and 0.95 mM, respectively (Table 3). Interestingly, both compounds

were more potent AGEs inhibitors as compared to the standard aminoguanidine ($IC_{50} = 1.8 \text{ mM}$). Therefore, from the above results, both alkaloids; (+)-neocadambine A (1) and (-)-nauclederine (2), can be potential lead compounds for the development of therapeutic agents for the prevention of AGEs formation.

TABLE 3. The IC₅₀ values of three different extracts of *N. cadamba*, (+)-neocadambine A (1), (-)-nauclederine (2) and aminoguanidine (standard)

Extracts/Compounds	IC ₅₀ (mg/mL)	IC ₅₀ (mM)
Hexane	> 1.0	-
Dichloromethane	0.15	-
Methanol	> 1.0	-
(+)-Neocadambine A (1)	-	1.2
(-)-Nauclederine (2)	-	0.95
Aminoguanidine (standard)	0.15	1.8

CONCLUSIONS

In conclusion, the current study has yielded two compounds; (+)-neocadambine A(1) and (-)-nauclederine (2) from the dichloromethane extract of the bark of N. cadamba which demonstrated high potency in inhibiting AGEs formation. (-)-Nauclederine (2) exhibited twice the potency of aminoguanidine which is a prototype therapeutic agent for the prevention of the formation of AGEs.

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REFERENCES

Ahmed, F., Rahman, S., Ahmed, N., Hossain, M., Biswas, A., Sarkar, S., Banna, H., Khatun, A., Chowdhury, M.H. & Rahmatullah, R. 2011. Evaluation of *Neolamarckia cadamba* (Roxb.) Bosser leaf extract on glucose tolerance in glucose-induced hyperglycemic mice. *African Journal of Traditional, Complementary, and Alternative Medicines* 8(1): 79-81.

Alam, M.A., Subhan, N., Chowdhury, S.A., Awal, M.A., Mostofa, M., Rashid, M.A., Hasan, M.H., Nahar, L. & Satyajit, D.S. 2011. Anthocephalus cadamba (Roxb.) Miq., Rubiaceae, extract shows hypoglycemic effect and eases oxidative stress in alloxan-induced diabetic rats. Revista Brasileira de Farmacognosia 21(1): 155-164.

Boisard, S., Ray, A.M.L., Gatto, J., Aumond, M.C., Blanchard, P., Derbré, S., Flurin, C. & Pascal, R. 2014. Chemical composition, antioxidant and anti-ages activities of a french poplar type propolis. *Journal of Agricultural and Food Chemistry* 62(6): 1344-1351.

Brownlee, M. 2001. Biochemistry and molecular cell biology of diabetic complications. *Nature* 414(6865): 813-820.

de Leeuw, K., Kallenberg, C. & Bijl, M. 2005. Accelerated atherosclerosis in patients with systemic autoimmune diseases. *Annals of the New York Academy of Sciences* 1051(1): 362-371.

DeGroot, J. 2004. The AGE of the matrix: Chemistry, consequence and cure. *Current Opinion in Pharmacology* 4(3): 301-305.

Derbré, S., Gatto, J., Pelleray, A., Coulon, L., Séraphin, D. & Richomme, P. 2010. Automating a 96-well microtiter plate assay for identification of AGEs inhibitors or inducers: Application to the screening of a small natural compounds library. *Analytical and Bioanalytical Chemistry* 398(4): 1747-1758.

Elosta, A., Ghous, T. & Ahmed, N. 2012. Natural products as anti-glycation agents: Possible therapeutic potential for diabetic complications. *Current Diabetes Reviews* 8(2): 92-108.

Grillo, M.A. & Colombatto, S. 2008. Advanced glycation end-products (AGEs): Involvement in aging and in neurodegenerative diseases. *Amino Acids* 35(1): 29-36.

- Habtemariam, S. 2019. Medicinal Foods as Potential Therapies for Type-2 Diabetes and Associated Diseases: Chemical and Pharmacological Evidences for Coffee as a Modulator of Type 2 Diabetes and Metabolic Syndrome. New York: Academic Press.
- Hu, X.J., Di, Y.T., Wang, Y.H., Kong, L.Y., Gao, S., Li, C.S., Liu, H.Y., He, H., Ding, J., Xie, H. & Hao, X. 2009. Carboline alkaloids from Trigonostemon lii. *Planta Medica* 75(10): 1157-1161.
- Jandeleit-Dahm, K. & Cooper, M.E. 2008. The role of AGEs in cardiovascular disease. *Current Pharmaceutical Design* 14(10): 979-986.
- Kurien, B.T., Hensley, K., Bachmann, M. & Scofield, R.H. 2006. Oxidatively modified autoantigens in autoimmune diseases. *Free Radical Biology and Medicine* 41(4): 549-556.
- Luevano-Contreras, C. & Chapman-Novakofski, K. 2010. Dietary advanced glycation end products and aging. *Nutrients* 2(12): 1247-1265.
- Martins, D. & Nunez, C.V. 2015. Secondary metabolites from Rubiaceae species. *Molecules* 20(7): 13422-13495.
- McLean, S., Dmitrienko, G.I. & Szakolcai, A. 1976. Constituents of Naucleadiderrichii. Part VII. Synthesis of nauclederine, naucleonine, and naucleonidine; spectroscopic evidence for the structures of 3α-dihydrocadambine and two other constituents. *Canadian Journal of Chemistry* 54(8): 1262-1277.
- Murray, D.G., Szakolcai, A. & McLean, S. 1972. The constituents of Naucleadiderrichii. Part III. Indole-pyridine alkaloids. *Canadian Journal of Chemistry* 50(10): 1486-1495.
- Pandey, A. & Negi, P.S. 2016. Traditional uses, phytochemistry and pharmacological properties of *Neolamarckia cadamba*: A review. *Journal of Ethnopharmacology* 181(1): 118-135.
- Peyroux, J. & Sternberg, M. 2006. Advanced glycation endproducts (AGEs): Pharmacological inhibition in diabetes. *Pathologie-Biologie* 54(7): 405-419.
- Poulsen, M.W., Hedegaard, R.V., Andersen, J.M., Courten, B.C.,
 Bügel, S., Nielsen, J., Skibsted, L.H. & Dragsted, L.O. 2013.
 Advanced glycation endproducts in food and their effects on health. Food and Chemical Toxicology 60: 10-37.
- Qureshi, A.K., Mukhtar, M.P., Hirasawa, Y., Hosoya, T., Nugroho, A.E., Morita, H., Shirota, O., Mohamad, K., Hadi, A.H.A., Litaudon, M. & Awang, K. 2011. Neolamarckines A and B, new indole alkaloids from *Neolamarckia cadamba*. *Chemical and Pharmaceutical Bulletin (Tokyo)* 59(2): 291-293.
- Reddy, V.P. & Beyaz, A. 2006. Inhibitors of the maillard reaction and age breakers as therapeutics for multiple diseases. *Drug Discovery Today* 11(13-14): 646-654.

- Séro, L., Sanguinet, L., Blanchard, P., Dang, B.T., Morel, M., Richomme, P., Séraphin, D. & Derbré, S. 2013. Tuning a 96-well microtiter plate fluorescence-based assay to identify AGE inhibitors in crude plant extracts. *Molecules (Basel, Switzerland)* 18(11): 14320-14339.
- Silva, D.H.S., Castro-Gamboa, L. & Bolzani, V.S. 2010. Plant diversity from Brazilian Cerrado and Atlantic Forest as a tool for prospecting potential therapeutic drugs. *Comprehensive Natural Products II* 3(1): 95-133.
- Singh, R., Barden, A., Mori, T. & Beilin, L. 2001. Advanced glycation end-products: A review. *Diabetologia* 44(2): 129-146.
- Sultana, N., Islam, M.T., Alencar, M.V.O.B.D., Silva, S.W.C., Chowdhury, M.U., Melo-Cavalcante, A.A.C. & Freitas, R.M.D. 2015. Phyto-pharmacological screenings of two Rubiaceae family plants. *African Journal of Pharmacy and Pharmacology* 9(31): 775-782.
- Takeuchi, M. & Yamagishi, S. 2008. Possible involvement of advanced glycation end-products (AGEs) in the pathogenesis of Alzheimer's disease. *Current Pharmaceutical Design* 14(10): 973-978.
- Thorpe, S.R. & Baynes, J.W. 1996. Role of the Maillard reaction in diabetes mellitus and diseases of aging. *Drugs Aging* 9(2): 69-77.
- Verma, R., Chaudhary, F. & Singh, A. 2018. Neolamarckia cadamba: A comprehensive pharmacological. Global Journal of Pharmacy & Pharmaceutical Sciences 6(4): 555691.
- Vlassara, H. & Uribarri, J. 2014. Advanced glycation end products (AGE) and diabetes: Cause, effect, or both? *Current Diabetes Reports* 14(1): 453-461.
- Wada, R. & Yagihashi, S. 2005. Role of advanced glycation end products and their receptors in development of diabetic neuropathy. *Annals of the New York Academy of Sciences* 1043(1): 598-604.
- Wang, B., Liu, L., Chen, Y.Y., Li, Q., Li, D., Liu, Y.P. & Luo, X.D. 2015. Monoterpenoid indole alkaloids from catharanthus roseus cultivated in Yunnan. *Natural Product Communications* 10(12): 1934578X1501001217.
- Wolffenbuttel, B.H., Giordano, D., Founds, H.W. & Bucala, R. 1996. Long-term assessment of glucose control by haemoglobin-AGE measurement. *Lancet* 347(9000): 513-515.
- Yuan, H.L., Zhao, Y.L., Qin, X.J., Liu, Y.P., Yu, H.F., Zhu, P.F., Jin, Q., Yang, X.W. & Luo, X.D. 2020. Anti-inflammatory and analgesic activities of *Neolamarckia cadamba* and its bioactive monoterpenoid indole alkaloids. *Journal of Ethnopharmacology* 260(1): 113103.

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