A FLUORESCENCE PHOSPHATE SENSOR BASED ON POLY(GLYCIDYL METHACRYLATE) MICROSPHERES WITH ALUMINIUM-MORIN

(Sensor Fosfat Berpendarfluor Berasaskan Mikrosfera Poli(Glisidil Metakrilat) dengan Aluminium-Morin)

Amalina Ahmad1, Norhadisah Mohd Zaini1, Normazida Rozi1, Nurul Huda Abd Karim1, Siti Aishah Hasbullah1, Lee Yook Heng1, Sharina Abu Hanifah1,2*

1School of Chemical Sciences and Food Technology
2Center for Water Research and Analysis
Faculty of Science and Technology,
Universiti Kebangsaan Malaysia, 43600 UKM Bangi, Selangor, Malaysia

*Corresponding author: sharina@ukm.edu.my

Received: 8 December 2015; Accepted: 11 March 2016

Abstract
The performance of new phosphate sensor was investigated using fluorescence spectrometer in the form of immobilized Al-morin on poly(glycidyl methacrylate) (pGMA) microspheres. pGMA microspheres that were synthesized by using suspension photopolymerization exhibited spherical-shaped morphology with diameters from 1.5 to 5.3 μm. The studies were carried out at pH 5 and the ratio of aluminium (III) chloride hexahydrate to morin was 3:1 (v/v). At pH 5, H2PO4− was measured at 548 nm emission wavelength. The relative fluorescence intensity was inversely proportional to H2PO4− concentrations. The linear range was observed between 6.6 – 58.8 μmol/L with detection limit (LOD) of 0.7 μmol/L. Ion interference study demonstrated that Al-morin was highly selective towards H2PO4−.

Keywords: phosphate sensor, polymer microspheres, aluminium-morin, fluorescence

Introduction
Phosphorus can be divided into organic phosphate and inorganic phosphate. Phosphate ions that move freely in water known as orthophosphate (P i) which is an inorganic phosphate [1]. High phosphate content in water can cause eutrophication. Eutrophication is the excess nutrients in aquatic ecosystems that bring to an excess algae growth and cause water and environmental problem [2]. Thus, monitoring phosphate in drinking water is vital to ensure that water quality follows the standard. The maximum concentration of phosphate recommended by World Health
Amalina et al: A FLUORESCENCE PHOSPHATE SENSOR BASED ON POLY(GLYCIDYL METHACRYLATE) MICROSPHERES WITH ALUMINIUM-MORIN

Organization (WHO) is $1 \text{mgL}^{-1}$[3]. Besides, the determination of phosphate concentrations in body fluids is necessary to diagnose hyperparathyroidism, hypertension [4], vitamin D deficiency, mineral and bone disorder, kidney failure [5] and Fractonia syndrome [6, 7].

Optical sensor is a device that converts light rays into electronic signals [8]. Fluorescence, luminescence, chemiluminescence and UV-visible spectrophotometers are commonly applied instruments in optical sensors [3]. Fluorescence method is widely used because it is highly sensitive, easy to operate, response rapidly and less costly [9]. Phosphate has known to quench aluminium-morin (Al-morin) fluorescence intensity. During 1950’s, Al-morin has been used to detect trace amount of fluoride ion and indirectly phosphate was determined to be an interfering ion [10]. Previously, Hong et al. [11] reported that Al-morin immobilized on PVA/PVC plasticized composite membrane based on sandwich configuration has been used in phosphate detection. This technique had improved the indicator leaching problem however produced narrow linear range of phosphate detection (11.0 – 51.4 µmol/L). In order to overcome its limitation, Lin et al. [12] claimed that Al-morin immobilized on pretreated PVC membrane is more sensitive as it produces wider linear range (44.1 – 110.2 µmol/L).

In this work, phosphate sensor was developed by immobilizing Al-morin onto poly(glycidyl methacrylate) (pGMA) microspheres which were synthesized by suspension photopolymerization technique. High surface area of pGMA microspheres is suitable for Al-morin immobilization matrix in order to increase the reaction site for phosphate detection. The microspheres are also a water-insoluble polymer, causing it to be applicable in phosphate sensing. All characterizations of immobilized Al-morin phosphate sensors were analyzed by fluorimetric method.

Materials and Methods

**Apparatus and reagent**
Chemicals including morin hydrate (98%, MP Biomedicals LLC), aluminium (III) chloride hexahydrate (AlCl$_3$.6H$_2$O) (99 %, Systerm), glycidyl methacrylate (97 %, Sigma Aldrich), ethylene glycol dimethacrylate (98 %, Sigma Aldrich), polyvinyl alcohol (98 %, Sigma Aldrich), 2,2-dimethoxy-2-phenylacetophenone (99 %, Sigma Aldrich), potassium dihydrogen phosphate (99 %, Systerm), potassium carbonate (K$_2$CO$_3$) (99 %, Sigma Aldrich), potassium acetate (CH$_3$COOK) (99 %, Sigma Aldrich), potassium chloride (KCl) (99 %, Sigma Aldrich), potassium fluoride (KF) (99 %, Sigma Aldrich), potassium nitrate (KNO$_3$) (99 %, BDH Chemicals Ltd), potassium sulfate (K$_2$SO$_4$) (99 %, Sigma Aldrich) and ethanol (95 %, Sigma Aldrich) were used as received.

The surface of microspheres was observed using Zeiss LEO 1450VP Scanning Electron Microscope (SEM). Infrared spectra of the microspheres were recorded by Fourier Transform Infrared spectrometer (Perkin Elmer) using attenuated total reflection (ATR-FTIR) spectrometry method. Response of Al-morin complex on fluorescence intensity was measured by Perkin Elmer Fluorescence Spectrometer at 548 nm emission wavelength.

**Synthesis of Al-morin complex**
A concentration of 23.2 µmol/L Morin solution was prepared in ethanol and 16.6 µmol/L aluminium (III) chloride hexahydrate (AlCl$_3$.6H$_2$O) in deionized water. Al-morin complex was formed by mixing AlCl$_3$.6H$_2$O with Morin in 3:1 (v/v). The formation of Al-morin was then confirmed when the fluorescence emission of Al-morin was determined to be at wavelength 510 nm as reported in the literature [12].

**Synthesis of poly(glycidyl methacrylate) (pGMA) microspheres**
An mount of 1 ml of glycidyl methacrylate (GMA), 1 ml of ethylene glycol dimethacrylate (EGDMA), 5 ml of 1% polyvinyl alcohol (PVA) and 1.6 % (w/w) of 2,2-dimethoxy-2-phenylacetophenone (DMPP) were placed in a vial. The mixture was sonicated within 15 minutes. The suspension was transferred into a petri dish and photocured under continuous nitrogen flow for 10 minutes. The microspheres were collected by centrifugation at 2000 rpm for 10 minutes. They were then left to dry at room temperature. pGMA microspheres were characterized by Scanning Electron Microscope (SEM) and Attenuated Total Reflectance-Fourier-transform Infrared (ATR-FTIR) spectrometer.
Immobilization of Al-morin onto pGMA microspheres
An amount of 20 – 30% (v/v) of Aluminium-Morin (Al-morin) complex was added into the mixture of GMA monomer, 1 ml ethylene glycol dimethacrylate (EGDMA), 5 ml of 1% polyvinyl alcohol (PVA) and 1.6% (w/w) of 2,2-dimethoxy-2-phenylacetophenone (DMPP) in a vial. The mixture was sonicated within 15 minutes then it was transferred into a petri dish for photocuring procedure under continuous nitrogen flow for 10 minutes. The microspheres were collected by centrifugation at 2000 rpm for 10 minutes and dried at room temperature. pGMA/Al-morin microspheres were characterized by SEM and ATR-FTIR.

pH analysis
The optimum pH for Al-morin to react with 110.2 µmol/L dihydrogen phosphate (H$_2$PO$_4$) was studied. pGMA/Al-morin microspheres were soaked in pH 1 buffer solution before they were next soaked into phosphate solution prepared at pH 1 for 5 minutes as reported previously [13]. These steps were repeated for analyzing the same concentration of phosphate solution at pHs ranging from 1 to 7. Fluorescence intensity before and after Al-morin reacts with phosphate were recorded by calculating the relative of fluorescence intensity at 548 nm.

Effect of phosphate concentration
pGMA/Al-morin microspheres were soaked in dihydrogen phosphate (H$_2$PO$_4$) solution prepared at pH 5 buffer with concentrations ranging from 0.7 – 124.9 µmol/L for 5 minutes. They were initially soaked at pH 5 buffer to remove the unreacted monomer. The fluorescence intensity of immobilized Al-morin before and after reacting with phosphate for each concentration was measured at 548 nm.

Effect of interference ions
Interference study was carried out by preparing a fixed concentration of H$_2$PO$_4$ (110.2 µmol/L) in the presence of common anions including carbonate (CO$_3^{2-}$), acetate (CH$_3$COO$^-$), chloride (Cl$^-$), fluoride (F$^-$), nitrate (NO$_3^-$) and sulfate (SO$_4^{2-}$). Each anion at the same concentration as H$_2$PO$_4$ was prepared. Fluorescence intensity of pGMA/Al-morin microspheres soaked in the analyte containing H$_2$PO$_4$ and interference ions were recorded at 548 nm.

Results and Discussion
Characterization of Al-morin immobilized onto pGMA
Morphology of pGMA microspheres
pGMA microspheres were used as a matrix to immobilize Al-morin complex. High surface area of these microspheres makes it possible to act as a matrix with good physical properties and these microspheres are chemically stable. pGMA microspheres were prepared using a rapid synthesis method, which is suspension photopolymerization. The droplets of monomer mixtures form pGMA at room temperature in the presence of DMPP. The polymerization process was terminated by the removal of the ultraviolet (UV) light source. The schematic polymerization of GMA is presented in Figure 1.
The spheres produced by this approach varied in size from 1.5 to 5.3 µm and are polydisperse. The size maintains even after Al-morin was immobilized onto pGMA microspheres. This observation might be due to the entrapment of chemical doping technique used to immobilize Al-morin which did not involve any chemical bonding. The presence of Al-morin causes pGMA/Al-morin microspheres to exhibit rougher surface compared to pGMA. The roughness of the microspheres surface has provides higher surface area [14]. SEM micrographs of pGMA microspheres and pGMA/Al-morin are presented in Figure 2.

Figure 2. Surface morphology of microspheres (A) pGMA and (B) pGMA with Al-morin

Spherical matrix is capable to maximize the surface area of sensor reagent and reduce the response time by allowing the analyte to penetrate through the matrix [15]. This shows that pGMA provides high surface area for phosphate detection and increases the chemisensor sensitivity. Microspheres optical chemisensor can reduce sample volume, increase sensitivity, shorten response time and reduce detection limit [16].

**ATR-FTIR spectra of pGMA/Al-morin**

ATR-FTIR spectra of pGMA and pGMA/Al-morin microspheres are shown in Figure 3. Both types of pGMA spectra have recorded a strong band at 1721 cm\(^{-1}\) (pGMA) and 1722 cm\(^{-1}\) (pGMA/Al-morin) due to C=O vibrations. Furthermore, bands of 843 cm\(^{-1}\) and 905 cm\(^{-1}\) correspond to the epoxy groups. No additional peak can be seen between pGMA and pGMA/Al-morin bands. This result confirms that the pGMA and Al-morin involved entrapment method which does not involve any chemical bonding between Al-morin complex and the GMA polymer.

**Optimization of pGMA/Al-morin**

**pH analysis**

Figures 4 shows graph of fluorescence intensity versus pH that was measured at 548 nm respectively. pH 5 was determined to be the optimum pH for pGMA/Al-morin to react with H\(_2\)PO\(_4\)\(^-\). Mohr and Wolfbeis [17] have reported that H\(_2\)PO\(_4\)\(^-\) is the predominant form of phosphate at pH 5. At pH <5, low fluorescence intensity could be related to the protonation of H\(_3\)PO\(_4\) to form H\(_2\)PO\(_4\) resulting in a poor ability of H\(_2\)PO\(_4\) to interact with Al-morin. Furthermore at pH >5, gradual decrease in fluorescence intensity is observed which may be due to hydroxide ions interference and formation of other form of phosphate ions [18].

**Effect of phosphate concentrations**

Morin (2',3,4',5,7-pentahydroxyflavone) is brown in colour when dissolves in ethanol and turn to light yellow solution as it is diluted. Free morin is weakly fluorescent and it becomes stronger as it forms a complexe with aluminium [19]. However, the fluorescence of Al-morin complex quenches distinctly when phosphate is introduced into the system [12]. The proposed reaction between Al-morin with H\(_2\)PO\(_4\)\(^-\) is shown in Figure 5.
Figure 3. ATR-FTIR spectra of (A) pGMA and (B) pGMA/Al-morin microspheres

Figure 4. Effect of pH on pGMA/Al-morin with $\text{H}_2\text{PO}_4^-$

Figure 5. Proposed chemical reaction of Al-morin with phosphate anion.

Study on the effect of phosphate concentration was carried out at pH 5 to investigate the ability of immobilized Al-morin in detecting $\text{H}_2\text{PO}_4^-$ at various concentrations. The pGMA/Al-morin microspheres was allowed to react with
H$_3$PO$_4^-$ at a fixed concentration within 5 minutes before analyzing by fluorescence spectrometer. Figure 6 shows the fluorescence intensity of pGMA/Al-morin reached equilibrium at 124.9 µmol/L H$_3$PO$_4^-$ with the linearity from 6.6 to 58.8 µmol/L (inset) and a detection limit as low as 0.7 µmol/L. This sensor produced higher range of phosphate concentration compared to poly(vinyl chloride) and poly(vinyl alcohol) based membrane may be due to larger surface area provided by pGMA microspheres [11, 12, 20].

The relative fluorescence intensity is inversely proportional to H$_3$PO$_4^-$ concentrations. The quenching of fluorescence could be further explained in Figure 7 [21]. It is possibly an example of CHEF (Chelation-Enhanced Fluorescence) and PET (Photoinduced Electron Transfer) phenomenons. CHEF effect is related to the photoinduced electron transfer (PET) mechanism. In the PET mechanism, exciting radiation induces electrons in the free ligand to transfer from the lone pairs on donor atoms (O-donors) to the π-system of the fluorophores and resulting in fluorescence quenching. These same lone pairs which involve in bonds formation with metal ions reduce the PET quenching effect. This occurrence leads to the CHEF effect, where metal ions can be detected by the increment of fluorescence intensity [22]. In this case, introducing H$_3$PO$_4^-$ to the system reduces the fluorescence intensity as aluminium detaches from morin by forming aluminium dihydrogenphosphate, Al(H$_2$PO$_4$)$_3$. This is due to poor fluorescence of non-chelating morin. Simultaneously, PET involves explaining about this phenomenon. Lone pairs
on morin O-donor atoms are no longer chelated with metal ion thus leading to increase PET quenching effect and indirectly reduce the fluorescence intensity.

**Effect of Ions Interference**

According to Table 1, pGMA/Al-morin was highly selective for $\text{H}_3\text{PO}_4^-$ over other anions which were commonly present in water samples such as $\text{CO}_3^{2-}$, $\text{CH}_3\text{COO}^-$, $\text{Cl}^-$, $\text{F}^-$, $\text{NO}_3^-$ and $\text{SO}_4^{2-}$. The higher affinity for $\text{H}_3\text{PO}_4^-$ over other anions may be caused by the presence of appropriately spaced cationic charges, Al$^{3+}$ [23]. All the interfering ions do not affect much on the detection of $\text{H}_3\text{PO}_4^-$ because the percentage of interference was less than 10% [24 – 26].

<table>
<thead>
<tr>
<th>Ion interference</th>
<th>Percentage of interference (%)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Carbonate, CO$_3^{2-}$</td>
<td>1.51</td>
</tr>
<tr>
<td>Acetate, CH$_3$COO$^-$</td>
<td>0.52</td>
</tr>
<tr>
<td>Chloride, Cl$^-$</td>
<td>5.43</td>
</tr>
<tr>
<td>Fluoride, F$^-$</td>
<td>0.00</td>
</tr>
<tr>
<td>Nitrate, NO$_3^-$</td>
<td>2.74</td>
</tr>
<tr>
<td>Sulfate, SO$_4^{2-}$</td>
<td>1.58</td>
</tr>
</tbody>
</table>

Performance of pGMA microspheres in the present study and PVC based membranes for immobilizing Al-morin is summarized in Table 2. It is clearly seen that pGMA/Al-morin has improved phosphate sensor performance compared to polyvinyl alcohol/ polyvinyl chloride/ aluminium-morin (PVA/PVC/Al-morin) composite membrane [11] and PVC/Al-morin matrix membrane [12] for $\text{H}_3\text{PO}_4^-$ detection. pGMA/Al-morin microspheres serves as a useful matrix for rapid phosphate measurement as the response time was recorded within 5 minutes.

<table>
<thead>
<tr>
<th>Parameters</th>
<th>pGMA/Al-morin microspheres</th>
<th>PVA/PVC/Al-morin membrane$^a$</th>
<th>PVC/Al-morin membrane$^b$</th>
</tr>
</thead>
<tbody>
<tr>
<td>pH</td>
<td>5</td>
<td>4</td>
<td>5</td>
</tr>
<tr>
<td>Dynamic range (µmol/L)</td>
<td>0.7 – 124.9</td>
<td>3.7 – 73.5</td>
<td>7.3 – 132.3</td>
</tr>
<tr>
<td>Linear range (µmol/L)</td>
<td>6.6 – 58.8</td>
<td>11.0 – 51.4</td>
<td>44.1 – 110.2</td>
</tr>
<tr>
<td>Limit of detection (µmol/L)</td>
<td>0.7</td>
<td>0.1</td>
<td>0.1</td>
</tr>
</tbody>
</table>


**Conclusion**

A convenient method for preparing pGMA/Al-morin microspheres for the development of optical phosphate sensor has been highlighted. pGMA/Al-morin microspheres have the ability to detect phosphate within a short time, high selectivity and able to measure phosphate concentrations at wider linear range caused by high surface area of the microspheres.
Acknowledgement

The authors would like to extend their gratitude towards Universiti Kebangsaan Malaysia for providing research facilities that was used in this research. This work was supported by the UKM grants DPP-2015-064 and ICONIC-2013-004.

References


